

An Outbreak of Lactose Fermenter Multidrug Resistant *Salmonella enterica serova typhi* in Sulaymani City, Iraq

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Abstract: In order to present the clinical microbiology characteristics of outbreaks of lactose fermenting *S. enterica serova typhi*, thirty lactose fermenting (L+) and three non-lactose fermenting (L-) *Salmonella enterica serova typhi* were isolated from blood of patients suffering from typhoid fever in indoor and outdoor of Sulaymani city-Iraq. The isolated *S. enterica* exhibited same characteristics as the type strain and L-*S. enterica* except for ONPG production. Serological examination revealed that all L+, L- and standard strain possessed the antigenic formula as *S. enterica*. The appearance of all L+*S. enterica* colonies on different media was similar to that of *E. coli*. The L+*S. enterica* isolates were 100% resist to Ampicillin, Cefalosporin, Chloramphenicol, Gentamycin, Tetracyclin and Trimethoprim antibiotics, While L-were sensitive to Amikacin, Cefalosporin, Cefotaxim, Ciprofloxacin, Chloramphenicol and Gentamycin. However 43.3% of L+*S. enterica* isolates were resist to all tested antibiotics, 6.7% were resist to 10 antibiotics out of eleven, 30% resist to nine antibiotics, 6.7% resist to 8 antibiotics, while 13.3 were resist to seven antibiotics.

Key words: Antibiotic resistant, *E. coli*, lactose fermenter, *Salmonella typhi*, sulaymani, typhoid fever

INTRODUCTION

Typhoid fever caused by *Salmonella enterica serovar typhi* remains endemic to many parts of Iraq, including Sulaymani city. To differentiate *Salmonella* from other *Enterobacteriaceae*, bacteriologists use lactose fermentation as a key biochemical test. As early as 1887, it was known that *Escherichia coli* was a lactose fermenter (L+) and that *Salmonella* was not a lactose fermenter (L-). Therefore, most differential plating media commonly developed and used today for the isolation of *Salmonella* contain lactose (Ewing *et al.*, 1986; Janda and Abbot, 1998). It has been reported that less than 1% of all *Salmonella* ferment lactose (Patrick *et al.*, 2000). There have been various reports of the occurrence of (Lac+) multidrug resist *Salmonella* in human (Chassy *et al.*, 1978; Corbion, 1981; Ezaki *et al.*, 1987; Camara *et al.*, 1989; Coovadia *et al.*, 1992; Patrick *et al.*, 2000; Eswarappa *et al.*, 2009). Until recently, the majority of *S. enterica* isolates from Sulaymani remained susceptible to Ciprofloxacin, Gentamycin and Amikacin, and only some cases of typhoid due to Lac+ multi-antibiotic resistant isolates of *S. enterica* have been documented, and therefore, these antibiotics still recommended as first line therapy for patients with typhoid fever.

Since 1907, there have been various reports of the occurrence of lactose fermenting *Salmonella* in humans, such as LacI *Salmonella enterica* serotype Virchow, *S. enterica* serotype Tennessee, *S. enterica* serotype

Indiana, *S. enterica* serotype Agona, *S. enterica* serotype Typhimurium, *S. enterica* serotype Oranienburg, *S. enterica* serotype Tuebingen, *S. enterica* serotype Newport, *S. enterica* serotype Typhi, *S. enterica* serotype Java, and *S. enterica* serotype Toulon (Camara *et al.*, 1989; Kukulka and Zimmer, 1989; Coovadia *et al.*, 1992; Louie *et al.*, 1993; Usera *et al.*, 1996; Usera *et al.*, 1998; Patrick *et al.*, 2000).

It is apparent that there are no reports that convey details of outbreaks caused by Lac+ multi-resistant strains of *S. enterica serova typhi* in Sulaymani city. The purpose of our work is to present the clinical microbiology characteristics of outbreaks of lactose fermenting *S. enterica serova typhi* in human blood suffering from typhoid fever in the middle of 2009 in indoor and outdoor patients in Sulaymani city, Iraq.

MATERIALS AND METHODS

Bacterial isolates: The isolates of *S. enterica* used in this study originated from blood of indoor and outdoor patients suffering from typhoid fever from General and Teaching Hospitals in the middle of 2009 in Sulaymani city Iraq.

Standard strain: The standard strain *Salmonella enterica* NCTC12023/ATCC14028 kindly provided by Media Diagnostic Center, Erbil, Iraq was used as reference strain in the study.

Table 1: Some biochemical tests of *S. enterica*

Bacteria	Sources of isolates	Oxidase	Kliglar (slope)	Kligler (bottom)	H ₂ S production	Minitol fermentation	Acid formation Xylose	Indolformati	Lactose fermentation
30 L+ <i>S. typhi</i>	Blood	-	Red alkaline	Acid yellow	Little H ₂ S	+	+	+	+
3 L- <i>S. typhi</i>	Blood	-	Red alkaline	Acid yellow		+	+	+	-
Standard strain	MCI	-	Red alkaline	Acid yellow		+	+	+	-

+: Positive; -: Negative

Bacteriology: Acid production from carbohydrates was tested in purple broth base, supplemented with 0.1 volume of a sterile 10% solution of each carbohydrate. The colonies were then screened for H₂S production and concurrently for lysine decarboxylase activity. Screening for H₂S production (black precipitate in the medium) was done with sulfide-indole-motility agar medium, and Kligler agar (Andrews and Hammack, 2000). In addition a commercial identification system API 20E system was used.

Serological examination test: Direct testing of colonies by slide agglutination tests were performed by using O-antisera, H and Vi antisera according to (Andrews and Hammack, 2000).

Standard bacterial strain *Salmonella enterica* NCTC12023/ATCC14028 was used as quality controls in these assays.

Susceptibility to antimicrobial agents: The susceptibility of the thirty lactose fermenting *S. enterica* isolates and three lactose-non fermenting colonial dissociate against eleven antimicrobial agents (Amikacin Ak, Ampicillin Amp, Cefalosporin Cef, Cefotaxim Cfm, Ciprofloxacin Cip, Chloramphenicol Chl, Gentamycin Gm, Rifampicin Rif, Streptomycin Str, Tetracyclin Tet and Trimethoprim Tri) was determined by means of disk diffusion method (Atlas *et al.*, 1995).

RESULTS

The thirty lactose-fermenting isolates (L+) and three non-lactose fermenting isolates (L-) exhibited the same characteristics as the type strain of *S. enterica* except for ONPG production, These L+ isolates were produced acid from lactose in purple broth base and were positive in O-nitrophenyl-beta-D- galactosidase (ONPG) test in the API 20E system. The L-isolates show an acid butt and alkaline slant on kligler (KI) agar with small quantity of H₂S visible in the Butt, these isolates produced no acid in purple broth base supplemented with lactose and gave a negative reaction in the ONPG test. Moreover the profile number obtained for API 20E test from the L+ isolates (7404543) were identified correctly. Standard strain was negative lactose fermented with the profile number of API 20 E test (6404540).

Acid from xylose for all L+, and L- isolate was positive Table 1. The lactose fermenting isolates consisted of motile, oxidase-negative, gram-negative rods. They produce colonies on MacConkey agar and turned triple

sugar iron agar yellow (Butt and slant) without gas formation. Indol production was observed on suitable media, these L+ isolates produced a small quantity of H₂S when grow on KI agar Table 1.

Serological examination revealed that all lactose fermenters, three non fermenting-lactose and standard strain possessed the antigenic formula of *S. enterica*. Thus, all lactose-fermenting isolates were identified as strains of *S. enterica serova typhi*, and the febrile illness was diagnosed as typhoid fever caused by lactose-fermenting *S. enterica serova typhi*.

The overall appearance of the Lac+ *S. typhi* colonies on different media was similar to that of *E. coli*, i.e., they appeared as rough, flat, lactose-fermenting bacterial colonies. All isolates recovered from plating on MacConky agar yielded rough, red colonies, and on Levine eosin-methylene blue agar, the colonies looked like small, rough, 2-4 mm green sequins with metallic sheen.

The L+ isolates were 100% resist to Amp, Cef, Chl, Gm, Tet and Tri antibiotics, whereas the L- dissociate was sensitive to, Ak, Cef, Cfm, Cip, Chl and Gm antibiotics Table 2.

DISCUSSION

Thirty isolates of lactose fermenting *S. enterica serova typhi* were isolated from blood of in patients and outpatients of teaching and general Hospitals, in Sulaymani city, Iraq. Lactose-fermenting *S. enterica* isolates obtained as the etiological agent of patients suffering from typhoid fever. All these isolates when identified were exhibited the same characteristics as the non-lactose fermented *S. enterica*, and standard strain except for β-galactosidase (ONPG) enzyme production, these colonies were similar to that of *E. coli*. The β-galactosidase enzyme produced by strains of Lac+*Salmonella* differs from *E. coli* β-galactosidase, offering further evidence that the operon did not originate from *E. coli* but could have originated from an enterobacterial ancestor common to *E. coli* (Cornelis, 1981; MacDonald and Riley, 1983; McClelland *et al.*, 2001). Bacteria belonging to the genus *Salmonella* are closely related to those belonging to the genus *Escherichia* and they have diverged from a common ancestor about 100 million years ago (Doolittle *et al.*, 1996). Despite their close relationship, *E. coli* has more than 800 genes that are absent in the *S. enterica* genome and more than 1, 100 *S. enterica* genes lack their homologues in *E. coli* (McClelland *et al.*, 2001). The

Table 2: Antibiogram of *S. enterica* isolated from blood of patient seafaring from typhoid fever

Antibiogram groups of L+ <i>S. typhi</i>	Isolates number	Antibiotics										
		Ak	Chl	Amp	Cef	Cfm	Rif	Gm	Cip	str	Tet	Tri
1	5,8,9,11,12,15,18,19,21,24,26,27,28	R	R	R	R	R	R	R	R	R	R	R
2	3,10	S	R	R	R	R	R	R	R	R	R	R
3	20,25	R	R	R	R	S	S	R	R	R	R	R
4	7,13,16,17,22,29,30	S	R	R	R	S	R	R	R	R	R	R
5	14,23	S	R	R	R	S	S	R	R	R	R	R
6	2	S	R	R	R	R	S	R	S	S	R	R
7	1,4,6	S	R	R	R	S	S	R	S	R	R	R
L- <i>S. typhi</i> isolates		S	S	R	S	S	R	S	S	R	R	R

R: Resist to antibiotics; S: Sensitive to antibiotics

region containing *Lac+* and *Lac* operon is one such locus and is present in *E. coli*, but absent in *S. enterica*. Thus *E. coli* is a lactose fermenter, whereas *S. enterica* is lactose non-fermenter. Nonetheless, diseases caused by lactose fermentation in extra-chromosomal genetic elements like plasmids (Patrick *et al.*, 2000; Eswarappa *et al.*, 2009), these elements can be horizontally transferred and acquired by bacteria (Hensel, 2004). The *Lac* operon consists of three genes, *lacZ*, *lac Y* and *lac A* which encode β -galactosidase (Wilson *et al.*, 2007). In this study and during the diagnosing it have been through that these isolates are resembling the *E. coli*.

Kohabata *et al.* (1983) thought that these L+ *S. enterica* isolates were to be identical to a strain 643 *Lac+* harboring the (*Lac+*) plasmid from ST-2 strain, strain 643 *Lac+* was derived from typical *S. enterica* 643 treated with the episome of strain ST-2 during the course of a genetic experiment. Thus, there have been some Salmonellosis reports of typhoid fever due to a naturally occurring *Lac+* *S. enterica* from different countries (LeMlnor *et al.*, 1974; Ananel *et al.*, 1980; Yanazaki and Kubota, 1982; Coovadia *et al.*, 1992; Patrick *et al.*, 2000; Wilson *et al.*, 2007).

The environment contain not only salmonellae but also other enteric bacteria, such as *Klebsiella pneumoniae* (MacDonald and Riley, 1983; Lucain and Piffaretti, 1983; Walia *et al.*, 1987), *Lactobacillus* (Chassy *et al.*, 1978), *Proteus* (Walia *et al.*, 1987), and *Serratia* (Walia *et al.*, 1987), which may carry the lactose operon on a plasmid or, in the case of some *Klebsiella* strains, on the chromosome (MacDonald and Riley, 1983). One theory for the origin of *Lac* in these plasmids involves the unlikely transfer of the operon from the chromosome of another enteric bacterium (*E. coli*) via a transposon (Cornelis, 1981) Some strains of *Klebsiella* carry the *lac* gene on the chromosome, there is evidence for may contain determinants of antimicrobial resistance, and there have been reports of outbreaks of *Lac+* and antimicrobial agent-resistant *Salmonella* in the literature (Timoney *et al.*, 1980; Threlfall *et al.*, 1983; Ezaki *et al.*, 1987). However 43.3% of our lactose fermenting *S. enterica* isolates were resist in vitro to all antimicrobials used, 6.7% were resist to 10 out of eleven antibiotics, 30% resist to nine antibiotics, 6.7% resist to 8 antibiotics, while 13.3 were resist to seven antibiotics Table 3. All L+S.

Table 3: Resistant of L+ *S. enterica* isolates to use antibiotics

% of L+ <i>S. typhi</i> isolates	Antibiotics
43.311	11
6.710	10
30.09	9
6.78	8
13.37	7

enterica isolates were 100% resist to Chl, Cef, Amp, Gm, Tet, and Tri, whereas the L-*S. enterica* isolates were sensitive to Ak, Cef, Chl, Cfm, Cip and Gm, and in some cases resistant to Chl. Multi-drug resist *S. enterica* which is reported in this study seems to confirm other reports of other workers (Nadgir *et al.*, 1998; Shrikala *et al.*, 1999; Sanghavi *et al.*, 1999; shrigala, 2004)

Cefalosporin, ciprofloxacin and Gentamycin has long been the known as the drug of choice for the treatment of typhoid fever in Sulaymani hospitals.

S. enterica is widely distributed in our environment and responsible for a wide range of clinical conditions some of them are fatal if untreated properly due to mistake in diagnosis specially typhoid fever and meningitis (Allen *et al.*, 2003). The early administration of antibiotic treatment has proven to be highly effective in eliminating infections, but indiscriminate use of antibiotics due to incorrect identification has led to the emergence of multidrug-resistant strains of *S. enterica* serovar Typhi.

After the cases due to a lactose-fermenting isolates occurred in August-October 2009, there has been no additional clinical cases of typhoid fever in indoor or outdoor patient were recorded in Sulaymani city, and the Ceftraixone was the main antibiotic for controlling typhoid fever in Sulaymani city.

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REFERENCES

- Allen, H.R., G.L. Alison, D.C. Harry and S. Maloy, 2003. Genetic rearrangements atrn operons in salmonella. Geneticsvol. 165: 95-959.

- Ananel, C.M., M.C. Finlayson, J.Z. Garson and M.L. Larson, 1980. A institutional outbreak of salmonellosis due to a lactose fermenting *Salmonella* new port. Am. J. Clin. Pathol., 74: 657-660.
- Andrews, W.H. and T. Hammack, 2000. Bacteriological analytical manual. FDA, US Food and Drug Administration.
- Atlas, R.M., L.C. Parks and A.E. Brown. 1995. Laboratory Manual of Experimental Microbiology. Mosby-Year Book, Inc., USA.
- Camara, F.P., M.A. Cardoso, de D.F. Ameida. 1989. Genetic analysis of lactose-fermenting *Salmonella typhimurium* isolated in Rio de Janeiro. Rev. Soc Bras Med Trop. 22: 81-83.
- Chassy, B.M., E.M. Gibson, A. Giuffrida, 1978. Evidence for plasmid-associated lactose metabolism in *Lactobacillus casei* sub sp. Casei. Curr. Microbiol., 1: 141-144.
- Corbion, B., 1981. Differential diagnosis of lactose fermenting *Salmonella* and certain enterobacteria. Bull. Inf. Lab. Serv. Vet., 81: 156-169.
- Cornelis, G. 1981. Sequence relationships between plasmids carrying genes for lactose utilization. J. Gen. Microbiol., 124: 91-97.
- Coovadia, Y.M, V. Gathiram, A. Bhamjee, R.M. Garratt, K. Mlisnaand and N. Pillay, 1992. A outbreak of Multiresistant *Salmonella typhi* in South Africa. QJM Int. J. Med., 82(2): 91-100.
- Doolittle, R.F., D.F. Feng, S. Tsang, G. Cho and E. Little, 1996. Determining divergence times of the major kingdoms of living organisms with a protein clock. Science, 271: 470-477.
- Eswarappa, S.M., G. Karnam, A.G. Nagarajan, S. Chakraborty and D. Chakravorty, 2009. Lac repressor is an virulence factor of *Salmonella enterica*: Its role in the evolution of virulence in salmonella. PLoS ONE, 4(6): e5789. doi: 10.1371/journal.pone.00057
- Ewing, W., H. Edwards and Ewings, 1986. Identification of the Enterobacteriaceae. Differentiation of Enterobacteriaceae by Biochemical Reactions. 4th Edn., Elsevier Science Publishing Co., New York, pp: 47-72.
- Ezaki, T., S.L. Liu, E. Yabuuchi, C. Sasakawa and M. Yoshikawa, 1987. Molecular characterization of a conjugative R-lac plasmid in *Salmonella typhi* isolated from a patient with typhoid fever. Ann. Inst. Pasteur. Microbiol., 138: 303-311.
- Hensel, M., 2004. Evolution of pathogenicity islands of *Salmonella enterica*. Int. J. Med. Microbiol., 294: 95-102.
- Janda, J.M. and S.L. Abbot, 1998. The Enterobacteria., Philadelphia, Pa: Lippincott-Raven Publishers. Historical perspectives on the family Enterobacteriaceae, in: pp: 1-12.
- Kukulska, D. and A. Zimmer, 1989. Selected properties of lactose-fermenting and non-fermenting *Salmonella agona* strains isolated from specimens from hospitalized infants. Med. Dosw. Mikrobiol., 41:115-120.
- Kohabata, S., M.T. akahashi and E. Yabuuchi, 1983. Lactose fermenting multiple drug-resistant *Salmonella typhi* strains isolated from a patient with post operative typhoid fever. J. Clin. Microbiol., 18(4): 920-925.
- LeMinor, L., C. Coynault and G. Pessoa, 1974. Determinisme plasmidique du caractere atypique (lactose positif) de souches de *S. typhimurium* et *S. oranienburg* isolees au Bresillors d'epidemies de 1971 a 1973. Ann. Microbiol. (Inst. Pasteur), 125A: 261-285.
- Louie, K.K., A.M. Paccagnella, W.D. sOsei, H. Lior, B.J. Francis and M.T. Osterholm, 1993. *Salmonella* serotype Tennessee in powdered milk products and infant formula-Canada and United States, 1993. Morbid. Mortal. Weekly Rep., 42: 516-517.
- Lucain, C. and J.C. Piffaretti, 1983. Characterization of a high Molecular mass resistance plasmid isolated from *Klebsiella pneumoniae* and coding for lactose degradation. FEMS Microbiol. Lett., 20: 131-134.
- MacDonald, C. and M. Riley, 1983. Cloning chromosomal lac genes of *Klebsiella pneumoniae*. Gene, 24: 341-345.
- McClelland, M., K.E. Sanderson, J. Spieth, S.W. Clifton and P. Latreille *et al.*, 2001. Complete genome sequence of *Salmonella enterica* serovar *typhimurium* LT2. Nature, 413: 852-856.
- Nadgir, S., B.V.S. Krishna, L.H. Halesh and S.S. Tallur, 1998. Multidrug resistant *Salmonella typhi* in Hubli. Ind. J. Med. Microbiol., 16(4): 185.
- Patrick, I.M., J.S. Sang and H.L. Lein, 2000. Diagnostic and public Health dilemma of lactose-fermenting *Salmonella enteric* serotype *typhimurium* in cattle in the northern United States. J. Clin. Microbiol., 38(3): 1221-1226.
- Sanghavi, S.K., M.P. Mane and K.B. Niphadkar, 1999. Multidrug drug resistant *Salmonella serotypes* Ind. J. Med. Microbiol., 17(2): 88-90.
- Shrikala, B., S. Shenoy, K. Vidyalakshmi and P. Pereira, 1999. Ciprofloxacin resistance in *Salmonella typhi*. Natl. Med. J. Ind., 12(3): 138.
- Shrigala, B., 2004. Drug resistance in *Salmonella typhi* Ojhas. 3(4): 1-2.
- Threlfall, E.J., M.L.M. Hall and B. Rowe, 1983. Lactose fermenting salmonellae in Britain. FEMS Microbiol. Lett., 17:127-130.
- Timoney, J.F., D.E. Taylor, S. Shin and P. McDonough, 1980. pJT2: Unusual H1 plasmid in a highly virulent lactose-positive and chloramphenicol-resistant *Salmonella typhimurium* strain from calves. Antimicrob. Agents Chemother., 18: 480-482.

- Usera, M.A., A. Echeita, A. Aladuená, M.C. Blanco, R. Reymundo, M.I. Prieto, O. Tello, R. Cano, D. Herrera and F. Martínez-Navarro, 1996. Interregional food borne salmonellosis outbreak due to powdered infant formula contaminated with lactose-fermenting *Salmonella virchow*. *Eur. J. Epidemiol.*, 12: 377–381.
- Usera, M.A., A. Rodríguez, A. Echeita and R. Cano, 1998. Multiple analysis of a food borne outbreak caused by infant formula contaminated by an atypical *Salmonella virchow* strain. *Eur. J. Clin. Microbiol. Infect. Dis.*, 17: 551–555.
- Walia, S.K., T. Madhavan, T.D. Chugh and K.B. Sharma, 1987. Characterization of self-transmissible plasmids determining lactose fermentation and multiple antibiotic resistance in clinical strains of *Klebsiella pneumoniae*. *Plasmid*, 17: 3-12.
- Wilson, C.J., H. Zhan, L. Swint-Kruse and K.S. Matthews, 2007. The lactose repressor system: Paradigms for regulation, allosteric behavior and protein folding. *Cell. Mol. Life Sci.*, 64: 3-16.
- Yanazaki, S. and H. Kubota, 1982. Atypical *Salmonella paratyphi*-B (d-tartrate+) strains from diarrhoeal stool of infants. *Media Circle*, 27: 333-342.