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Aerobic Bacterial Flora of Conjunctival Sac in the Healthy Iranian Water Buffalo (*Bubalus bubalis*)

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Abstract: This study was aimed to identify the normal aerobic bacterial flora of the conjunctiva in the Iranian water buffalo and to determine the effect of gender, age and ecotype variations on the flora. Fifty healthy Iranian water buffalos (33 females, 17 males), aged 1-15 years, of four different ecotypes-Azeri, Mazandarani, Khuzestani and Guilani were selected and divided into 4 age groups. Swabs were taken from the inferior conjunctival sac of both eyes (n = 100) and cultured on blood and MacConkey agar in aerobic condition. Forty seven buffalos out of 50 (94%) were positive for bacteria; 44/50 (88%) had positive culture from both eyes. The most frequent isolates were *Staphylococcus* sp., *Escherichia coli* and *Bacillus cereus*. Statistical analysis did not show significant difference in frequency of isolates between sexes, ecotypes and age groups (p>0.05). A correlation was found between the buffalo age and number of species isolated per eye (p<0.01). In this study *Dermatophilus congolensis* and *Edwardsiella tarda* were reported for the first time in the conjunctiva of animals. Gram-positive aerobes were most commonly cultured from the conjunctival sac of healthy water buffalos.

Keywords: Bacterial flora, conjunctiva, *Dermatophilus congolensis*, *Edwardsiella tarda*, water buffalo

INTRODUCTION

The microbial flora of the ocular surface consists of resident and transient bacterial, fungal (Andrew et al., 2003; Dupont et al., 1994), mycoplasmal (Dagnall, 1994; Barber et al., 1986; Rosendal, 1973; Campbell et al., 1973) and chlamydial (Pinard et al., 2002; Davidson et al., 1994) organisms that exist in a balance between themselves and immune system of the host. The resident microbial flora aids in maintaining ocular surface health by preventing overgrowth of potentially pathogenic agents. However in some circumstances (e.g., when surface defense mechanisms are weakened) this normal flora may become opportunists and cause infection (McClellan, 1997; Gilger, 2008). Therefore, characterization of normal flora of conjunctiva may be useful in treating eye infections. Bacterial flora of the normal conjunctiva has been reported in various animals (Moore and Nasisse, 1998). Gram-positive bacteria were reported as the most common isolates. However microbiota of the ocular surface depends on season of sampling, geographic location, housing conditions and the age of the animal (Ramsey, 1998).

Normal conjunctival bacterial flora has been reported in swamp buffalo (Tantivanich *et al.*, 1988) and bison (Davidson *et al.*, 1999), but to the authors' knowledge, conjunctival flora of healthy riverine buffalos has not been reported.

In Iran, almost half a million water buffaloes (river-type) are farmed, with 75% of its population concentrated in the northwest (Azeri ecotype), 18% in the southwest (Khuzestani ecotype) and 7% in the north of the country (Guilani and Mazandarani ecotypes) (Naserian and Saremi, 2007).

The purpose of this study was to identify the conjunctival aerobic bacterial flora and to determine the effect of gender, age and ecotype variations on this flora in the healthy Iranian water buffalo.

MATERIALS AND METHODS

This study was carried out in the summer of 2009 at the buffalo breeding and extension training center of Urmia, northwest of Iran. Fifty clinically normal Iranian water buffaloes (33 females, 66%; 17 males, 34%) with an age range of 1 to 15 years old in 4 age groups-yearlings 19 cases (3%), 2-5 years 8 cases (16%), 6-10 years 2 cases (44%) and 11-15 years 21 cases (42%) were used. The animals were of 4 Azeri (25 cases; 50%), Mazandarani (13 cases; 26%), Khuzestani (8 cases; 16%), and Guilani (4 cases; 8%) ecotypes. The buffalos were run through a specially designed chute. The eyes were examined by direct ophthalmoscopy to rule out the presence of any signs of ocular disease. Samples were taken from the inferior

conjunctival sac of both eyes using a sterile swab for each eye, avoiding swab contact with the eyelashes or skin of the eyelids. Swabs were placed in individual sterile test tubes and submitted immediately to the microbiology laboratory of center. In the laboratory swabs were cultivated on 5% ovine blood agar (Merck, Germany) and MacConkey agar (Himedia, India) and incubated at 37°C for 24-48 h, in aerobic condition. Different colony types were further isolated on blood agar plates. Colonies isolated from all plates were identified by using standard microbiological and biochemical methods (Quinn *et al.*, 1994).

Spearman's correlation test was used to measure of relationship the age of buffalo with number of bacterial species isolated per eye. ANOVA test was used to compare the number of isolates between ecotypes and age groups of buffalos and Student's t-test to compare the number of isolates between sexes using a software package (SPSS 16.0) with the statistical significance set at p<0.05.

RESULTS AND DISCUSSION

A total of 91 (91%) eyes were positive for bacteria. No bacteria were isolated in 3 cases (6 eyes; i.e., 6%). Single bacterial species were isolated in 12 (13.19%) eyes. Two different bacterial species were isolated in 54 (59.34%) eyes; 3 different bacterial species were isolated in 25 (27.47%) eyes. Based on Spearman's correlation test, there was a positive correlation between age of buffalo and number of the bacterial species isolated per eye (R = 0.449; p = 0.001).

The bacterial isolates consisted of 10 species belonging to 8 genera (Table 1). Gram-positive organisms were the predominant bacteria, representing 59% of isolates. Members of the genus Staphylococcus (29.20%) were the most prevalent. Genera of Dermatophilus and Streptococcus were less commonly isolated gram-positive organisms. Gram-negative organisms comprised 41% of isolates, with Escherichia coli being the most prevalent. Edwardsiella, Klebsiella and Proteus were infrequently isolated gram-negative genera. E. coli was the most frequent isolate in the 2-5 years group and Mazandarani ecotype, but in bothsexes, other age groups and ecotypes, Staphylococcus sp., were the predominant isolates. Difference in quantity of isolates between sexes, ecotypes and age groups was not significant (p>0.05).

Many studies have shown that gram-positive bacteria are usually predominant over gram-negative microorganisms in healthy eyes. This is consistent with our data where 59% of the isolates were gram-positive bacteria and *Staphylococcus* sp., predominated. Similar results have been found in healthy eye of horses (Vidal et al., 2010), sheep (Dagnall, 1994), dogs (Prado et al., 2005), cats (Espinola and Linelbaum, 1996), domestic rabbits (Cooper et al., 2001), llamas (Gionfriddo et al., 1991), Asian elephants (Tantivanich et al., 2002), mule deers (Dubay et al., 2000), raccoons (Spinelli et al., 2010), opossums (Pinard et al., 2002), iguanas (Taddei et al., 2010), birds (Dupont et al., 1994; Zenoble et al., 1983; Wolf et al., 1983) and in the swamp buffalos (Tantivanich et al., 1988).

Table 1: Aerobic bacteria recovered from conjunctival sac of clinically normal Iranian water buffalo (Bubalus bubalis)

	Number of Isolates (%)										
		Sex		Ecotype				Age group			
Bacteria	Total	1	2	3	4	5	6	7	8	9	10
Gram-positive											
Bacillus cereus	43 (22.05)	14 (7.18)	29 (14.87)	17 (8.72)	3 (1.54)	8 (4.10)	5 (2.56)	15 (7.69)	5 (2.56)	1 (0.51)	22 (11.28)
Staphylococcus epidermidis	36 (18.46)	11 (5.64)	25 (12.82)	15 (7.69)	4 (2.05)	11 (5.64)	6 (3.08)	12 (6.15)	7 (3.59)	2 (1.02)	15 (7.70)
Ŝtaphylococcus intermedius	17 (8.72)	6 (3.08)	11 (5.64)	7 (3.59)	1 (0.51)	3 (1.51)	6 (3.08)	4 (2.05)	2 (1.05)	1 (0.51)	10 (5.13)
Dermatophilus congolensis	14 (7.18)	7 (3.59)	7 (3.59)	11 (5.64)	1 (0.51)	1 (0.51)	1 (0.51)	8 (4.10)	3 (1.53)	1 (0.51)	2 (1.05)
Staphylococcus aureus	4 (2.05)	1 (0.51)	3 (1.53)	-	2 (1.05)	-	2 (1.05)	-	-	-	4 (2.05)
Streptococcus zooepidemicus Gram-negative	1 (0.51)	-	1 (0.51)	-	-	-	1 (0.51)	-	-	-	1 (0.51)
Escherichia coli	52 (26.67)	16 (8.2)	36 (18.47)	20 (10.25)	6 (3.10)	9 (4.61)	17 (8.71)	11 (5.64)	10 (5.12)	3 (1.54)	28 (14.37)
Edwardsiella tarda	14 (7.18)	5 (2.56)	9 (4.61)	5 (2.56)	2 (1.02)	4 (2.05)	3 (1.53)	3 (1.53)	2 (1.02)	1 (0.51)	8 (4.10)
Klebsiella spp	9 (4.61)	4 (2.05)	5 (2.56)	5 (2.56)	-	1 (0.51)	3 (1.53)	4 (2.05)	1 (0.51)	-	4 (2.05)
Proteus spp	5 (2.55)	2 (1.05)	3 (1.53)	3 (1.53)	-	- ` ′	2 (1.05)	3 (1.53)	- ` ′	-	2 (1.05)

^{1:} Male; 2: Female; 3: Azeri; 4: Guilani; 5: Khozestani; 6: Mazandarani; 7: yearlings; 8: 2-5 years of age; 9: 6-10 years of age; 10: 11-15 years of age

The most common gram-negative organisms present in the eyes vary among the species. *E.coli* was reported to be the most common gram-negative organism in pigs (Davidson *et al.*, 1994) and camels (Fahmi *et al.*, 2003), as well as in the present study in water buffalos.

D. congolensis with frequency of 7.18% is one of isolates that is reported for the first time from conjunctiva of animals in the present study. This organism is primarily known as a cutaneous pathogen in buffalos (Pal, 1995; Sharma et al., 1992) and other animals (Pal, 1995), but it is able to cause disease in other tissues of various animals, such as oral cavity in buffalos (Kharole et al., 1976) and cattle (Ibu et al., 1987), lymph node in goats (Singh and Murty, 1978) and subcutaneous tissue in cats (Cararostas et al., 1984). In Iran, there are reports of cutaneous form of disease in cattle (Jafari Shoorijeh et al., 2008) and sheep (Hashemi et al., 2004).

E. tarda with frequency of 7.18% was isolated from conjunctiva of animals for the first time in this study. This organism is principally associated with freshwater sources and animals that inhabit these ecosystems, such as amphibia, reptile and fish (Janda et al., 1991). The buffalo is also one of those animals that like aquatic environments. The infections associated with this organism include gasteroenteritis, septicemia, meningitis and wound infections such as cellulitis or gas gangrene associated with trauma to mucosal surfaces in humans (Nelson et al., 2009; Janda et al., 1991), septicemia, chronic enteritis and emphysematous putrefactive disease in animals (Galal et al., 2005; Meyer et al., 1973).

Considering the presence *D. congolensis* and *E. tarda* in ocular surface of buffalos, external eye infections with these opportunistic pathogens can be expected in situations where the normal defense mechanism of the ocular surface is damaged.

In our study diversity of nonpathogenic and potentially pathogenic bacteria was isolated, but, *Moraxella bovis* as a primary cause of infectious bovine keratoconjunctivitis (Brown *et al.*, 1998) was not isolated. This agent has been reported from healthy eyes of cattle (Powe *et al.*, 1992; Barber *et al.*, 1986; Wilcox, 1970) and bison (Davidson *et al.*, 1999), but not in swamp buffalo (Tantivanich *et al.*, 1988). In addition, this potentially pathogen has not been isolated from diseased eye of riverine buffalos (Rajesh *et al.*, 2009). It seems that *M. bovis* is not a common isolate of ocular surface in both affected and unaffected water buffalos.

It had previously been confirmed that age has various effects on the conjunctival normal flora in humans (Liu *et al.*, 2011; Rubio, 2006) and animals

(Andrew *et al.*, 2003; Cooper *et al.*, 2001; Whitley, 2000). Liu *et al.* (2011) gave evidence that age has a significant correlation with number of species per eye in humans (Liu *et al.*, 2011). Our study produced similar results to Liu *et al.* (2011), as the number of species of conjunctival bacterial flora from the older buffalos was greater than that of younger. This may be due to aged buffalos having a lower resistance from depressed ocular defense mechanisms (McClellan, 1997).

CONCLUSION

The results of this study were in agreement with previous studies indicating *Staphylococcus* spp as the most common inhabitant of the eyes of most animal species. This is first report of presence of *D. congolensis* and *E. tarda*, pathogenic and opportunist organisms, in the normal conjunctiva of animals. Incidence of bacterial species of conjunctival flora can be influenced by age of buffalo.

REFERENCES

- Andrew, S.E., A. Nguyen, G.L. Jones and D.E. Brooks, 2003. Seasonal effects on the aerobic bacterial and fungal conjunctival flora of normal thoroughbred brood mares in florida. Vet. Ophthalmol., 6(1): 45-50.
- Barber, D.M.L., G.E. Jones and A. Wood, 1986. Microbial flora of the eyes of cattle. Vet. Rec., 118(8): 204-206.
- Brown, M.H., A.H. Brightman, B.W. Fenwick and M.A. Rider, 1998. Infectious bovine keratoconjunctivitis: A review. J. Vet. Internal. Med., 12(4): 259-266.
- Campbell, L.H., J.G. Fox and S.B. Snyder, 1973. Ocular bacteria and mycoplasma of the clinically normal cat. Feline Pract., 3(10): 10-12.
- Cararostas, M.C., R. Miller and M.G. Woodward, 1984. Subcutaneous dermatophilosis in a cat. J. Am. Vet. Med. Assoc., 185(6): 675-676.
- Cooper, S.C., G.J. McLellan and A.N. Rycroft, 2001. Conjunctival flora observed in 70 healthy domestic rabbits (*Oryctolagus cuniculus*). Vet. Rec., 149(8): 232-235.
- Dagnall, G.J.R., 1994. A investigation of colonization of the conjunctival sac of sheep by bacteria and mycoplasmas. Epidemiol. Infect., 112(3): 561-567.
- Davidson, H.J., D.P. Rogers, T.J. Yeary, G.G. Stone,D.A. Schoneweis and M.M. Chengappa, 1994.Conjunctival microbial flora of clinically normal pigs. Am. J. Vet. Res., 55(7): 949-951.

- Davidson, H.J., J.G. Vestweber, A.H. Brightman, T.H.V. Slyke, L.K. Cox and M.M. Chengappa, 1999. Ophthalmic examination and conjunctival bacteriologic culture results from a herd of North America bison. J. Am. Vet. Med. Assoc., 215(8): 1142-1144.
- Dubay, S.A., E.S. Williams, K. Mills and A.M. Boerger-Fields, 2000. Bacterial and nematodes in the conjunctiva of mule deer from Wyoming and Utah. J. Wild Life. Dis., 36(4): 783-787.
- Dupont, C., M. Carrier and R. Higgins, 1994. Bacterial and fungal flora in healthy eyes of birds of prey. Can Vet. J., 35(11): 699-671.
- Espinola, M.B. and W. Lilenbaum, 1996. Prevalence of bacteria in the conjunctival sac and on the eyelid margin of clinically normal cats. J. Small Anim. Pract., 37(8): 364-366.
- Fahmi, L.S., A.A. Hegazy, M.A. Abdelhamid, M.E. Hatem and A.A. Shamaa, 2003. Studies on eye affections among camels in Egypt: Clinical and bacteriological studies. Sci. J. King Faisal Univ., 4(2): 159-176.
- Galal, N.F., S.M. Ismail, R.H. Khalil and M.K. Soliman, 2005. Studies on edwardsiella infection in oreochromis niloticus. Egypt J. Aqua. Res., 31(1): 460-471.
- Gilger, B.C., 2008. Immunology of the ocular surface. Vet. Clin. North Am. Small Anim. Pract., 38(2): 223-231.
- Gionfriddo, J.R., R. Rosenbusch, J.M. Kinyon, D.M. Betts and T.M. Smith, 1991. Bacterial and mycoplasmal flora of the healthy camelid conjunctival sac. Am. J. Vet. Res., 52(7): 1061-1064.
- Hashemi, T.G.R., M. Rad and M. Chavoshi, 2004. A survey on dermatophilosis in sheep in the north of Iran. Iran J. Vet. Res., 5(2): 97-101.
- Ibu, J., A.A. Makinde and U.R. Nawath, 1987. Oral dermatophilosis in imported cattle in Nigeria. Vet. Rec., 120(2): 42.
- Jafari, S.S., K.H. Badiee, M.A. Behzadi and A. Tamadon, 2008. First report of dermatophilus congolensis dermatitis in dairy cows in Shiraz, Southern Iran. Iran J. Vet. Res., 9(3): 281-283.
- Janda, J.M., S.L. Abbott, S. Kroske-Bystrom, K.W. Wendy, W.K.W. Cheung, C. Powers, R.P. Kokka and K. Tamura, 1991. Pathogenic properties of edwardsiella species. J. Clin. Microbiol., 29(9): 1997-2001.
- Kharole, M.U., P.P. Gupta, B. Singh and P.N. Dhingra, 1976. Dermatophilosis (streptothricosis) in a buffalo calf (bubalus bubalis). Zentbl. Vet. Med. B., 23(7): 604-608.

- Liu, J., J. Li, J. Huo and H. Xie, 2011. Identification and quantitation of conjunctival aerobic bacterial flora from healthy residents at different ages in Southwest China. Afr. J. Microbiol. Res., 5(3): 192-197.
- McClellan, K.A., 1997. Mucosal defense of the outer eye. Surv. Ophthalmol., 42(3): 233-246.
- Meyer, F.P. and G.L. Bullock, 1973. Edwardsiella tarda, a new pathogen of channel catfish (ictalurus punctatus). Appl. Microbiol., 25(1): 155-156.
- Moore, C.P. and M.P. Nasisse, 1998. Clinical Microbiology. In: Gelatt, K.N. (Ed.), Veterinary Ophthalmology. Lea and Febiger, Philadelphia, pp: 259-289.
- Naserian, A.A and B. Saremi, 2007. Water buffalo industry in Iran. Ital. J. Anim. Sci., 6(Suppl 2): 1404-1405.
- Nelson, J.J., C.A. Nelson and J.E. Carter, 2009. Extraintestinal manifestations of edwardsiella J. La. State. Med. Soc., 161(2): 103-106.
- Pal, M., 1995. Prevalence in India of dermatophilus congolensis infection in clinical specimens from animals and humans. Rev. Sci. Tech. Off. Int. Epiz., 14(3): 857-863.
- Pinard, C.L., A.H. Brightman, T.J. Yeary, T.D. Everson, L.K. Cox, M.M. Chengappa and H.J. Davidson, 2002. Normal conjunctival flora in the North American opossum (didelphis virginiana) and Raccoon (Procyon lotor). J. Wildlife. Dis., 38(4): 851-855.
- Powe, T.A., K.E. Nusbaum, T.R. Hoover, S.R. Rossmanith and P.C. Smith, 1992. Prevanlence of nonclinical Moraxella bovis infections in bulls as determined by ocular culture and serum antibody titer. J. Vet. Diagn. Invest., 4(1): 78-79.
- Prado, M.R., M.F.G. Rocha, E.H.S. Brito, M.D. Girao, A.J. Monteiro, M.F.S. Teixeira and J.J.C. Sidrim, 2005. Survey of bacterial microorganisms in the conjunctival sac of clinically normal dogs and dogs with ulcerative keratitis in Fortaleza, Ceara, Brazil. Vet. Ophthalmol., 8(1): 33-37.
- Quinn, P.J., M.E. Carter, B.K. Markey and G.R. Carter, 1994. Clinical Veterinary Microbiology. Wolf, London.
- Rajesh, K., K. Suresh and N.S. Sundar, 2009.Infectious bovine keratoconjunctivitis in a buffalo-clinical and therapeutic aspects. Buffalo Bull, 28(3): 110-112.
- Ramsey, D.T., 1998. Surface Ocular Microbiology in Food and Fiber-Producing Animal. In: Howard, J.L. and R.A. Smith (Eds.), Current Veterinary Therapy 4, Food Animal Practice. Sunders, Philadelphia, pp. 648-649.

- Rosendal, S., 1973. Canine mycoplasmas I: Cultivation from conjunctiva, respiratory and genital tracts. Acta. Patholog. Microbio., 81(4): 441-445.
- Rubio, E.F., 2006. Influence of age on conjunctival bacteria of patients undergoing cataract surgery. J. Eye., 20(4): 447-454.
- Sharma, R.D., M.S. Kwatra and S.S. Dhillon, 1992. Dermatophilosis outbreak in buffaloes in Punjab. Buffalo J., 3(2): 293-296.
- Singh, V.P. and D.K. Murty, 1978. An outbreak of dermatophilus congolensis infection in goats. Ind. Vet. J., 55(3): 674-676.
- Spinelli, T.P., E.F. Oliveira-Filho, D. Silva, R. Mota and F.B. Sa, 2010. Normal aerobic bacterial conjunctival flora in the crab-eating raccoon (procyon cancrivorus) and coati (nasua nasua) housed in captivity in pernambuco and paraiba (Northeast, Brazil). Vet. Ophthalmol., 13(Suppl1): 134-136.
- Taddei, S., P.L. Dodi, F.D. Lanni, C.S. Cabassi and S. Cavirani, 2010. Conjunctival flora of clinically normal captive green iguanas (iguana iguana). Vet. Rec., 167(1): 29-30.
- Tantivanich, P., P. Suttipong and O. Naweparp, 1988. Conjunctival flora of normal Thai swamp buffalo. Thai J. Vet. Med., 18(2): 159-165.

- Tantivanich, P., K. Soontornvipart, N. Tuntivanich, S. Wongaummuaykul and P. Brikawan, 2002. Conjunctival microflora in clinically normal Asian elephants in Thailand. Vet. Res. Commun., 26(4): 251-254.
- Vidal, G.H., R.R. Romero, L.E.R. Tovar, F.A.M. Valdez and J.A.V. Contreras, 2010. Localization of serratia marcescens in bacterial and fungal profile of conjunctiva of clinically healthy horses from monterrey, nuevo leon, Mexico. Vet. Mex., 41(4): 239-249.
- Whitley, R.D., 2000. Canine and feline primary ocular bacterial infections. Vet. Clin. North Am. Small Anim. Pact, 30(5): 1151-1167.
- Wilcox, G.E., 1970. Bacterial flora of the bovine eye with special reference to the Moraxella and Neisseria. Aust. Vet. J., 46(6): 253-257.
- Wolf, E.D., K. Amass and J. Olsen, 1983. Survey of conjunctival flora in the eye of clinically normal captive exotic birds. J. Am. Vet. Med. Assoc., 183(11): 1232-1233.
- Zenoble, R.D., R.W. Griffith and S.L. Clubb, 1983. Survey of bacteriologic flora of conjunctiva and cornea in healthy psittacine birds. Am. J. Vet. Res., 44(10): 1966-1967.